The Expression Profile Evaluation of Mir-125b in Tuberculosis and Non-Small Cell Lung Cancer Patients

Abstract

Background and Aim: Worldwide, lung cancer, specifically nonsmall-cell lung cancer (NSCLC), is one of the most preventable and common cancer types. Tuberculosis (TB) is a major global public health threat, and the most important infectious disease throughout history in the world. Having TB before lung cancer significantly correlates with mortality due to lung cancer. TB and lung cancer have similar symptoms, and TB has symptoms identical to malignancy, so sometimes its diagnosis is confused with lung cancer. The miRNAs are biological molecules that play critical regulatory roles in the physiological and pathological processes. In several research types, many miRNAs have been studied, which showed the alternation of expression in TB and NSCLC separately. In this research paper, the expression of mir-125b was examined, which is commonly altered in TB and NSCLC. Materials and Methods: Thirty patients with NSCLC, thirty patients with TB who were new cases, and thirty healthy individuals were contributed to this research. The expression pattern of mir-125b was evaluated and compared in TB and NSCLC patients with healthy controls, using the real-time polymerase chain reaction method. Results: As a result, the expression of mir-125b was lower in TB and NSCLC patients than in healthy controls. Conclusion: Therefore, mir-125b can be used as a prognostic biomarker in TB and NSCLC diseases. The expression pattern of mir-125b could be useful to follow-up on the biological pathways that have essential roles in these diseases.

Keywords: Expression profile, mir-125b, nonsmall-cell lung cancer, tuberculosis

Introduction

Lung cancer is the second most common cancer among men and is one of the most preventable types of cancer. The types of lung cancer are classified based on the appearance of the cells under the microscope. Nonsmall-cell lung cancer (NSCLC) is also divided into three categories: (1) superficial tissue cancer, (2) mucosal epithelial gland cancer (glandular epithelium), and (3) large-cell lung cancer. Among those with cancer, about 85% are NSCLC, and about 15% are SCLC.^[1] Worldwide, lung cancer is the most common cancer in prevalence and mortality in Europe and North America. About 85% of lung cancer cases are associated with smoking. About 40% of lung cancers are "adenocarcinomas," which usually form in the lateral lung tissue. Sclerotic cell carcinoma accounts for about 30% of all lung cancers, and about 9% are "large-cell carcinoma.^[2]

Tuberculosis (TB) disease is a major global public health threat^[3] and is one of the most

For reprints contact: WKHLRPMedknow_reprints@wolterskluwer.com

important infectious diseases throughout history in the world. Various communities are continually trying to control and combat this disease. One-third of the world population is latently infected with *Mycobacterium tuberculosis*, a notorious intracellular pathogen, with around 1.6 million deaths recorded in 2017.

MicroRNAs are small evolutionarily conserved nucleotide sequences that regulate gene expression after transcription, by mRNA degradation or inhibition of translation.^[4] MicroRNAs are involved in many transformative biological processes proliferation. such as development, apoptosis,^[5] metabolism, stem cell differentiation, and disease development.^[6,7] The aberrant miRNAs expression is also involved in many human diseases and disorders including cancer,^[8,9] diabetes, kidney diseases. neurodegenerative diseases, liver diseases, heart diseases, and altered immune system function.[10,11]

Mir-125b is one of the miRNAs which shows expression changes in TB and NSCLC. Some researchers have shown

How to cite this article: Shahsavani M, Baghbani-Arani F, Sheikhpour M. The expression profile evaluation of Mir-125b in tuberculosis and Non-Small Cell lung cancer patients. Clin Cancer Investig J 2021;10:60-4.

Mahboubeh Shahsavani¹, Fahimeh Baghbani-Arani¹, Mojgan Sheikhpour^{2,3}

¹Department of Genetics and Biotechnology, School of Biological Science, Varamin-Pishva Branch, Islamic Azad University, Varamin, ²Department of Mycobacteriology and Pulmonary Research, Pasteur Institute of Iran, ³Microbiology Research Center, Pasteur Institute of Iran, Tehran, Iran

Submitted: 18-Dec-2020 Revised: 30-Jan-2021 Accepted: 01-Mar-2021 Published: 23-Apr-2021

Address for correspondence: Dr. Fahimeh Baghbani-Arani, Department of Genetics and Biotechnology, School of Biological Science, Islamic Azad University, Varamin-Pishva Branch, Varamin, Iran. E-mail: baghbani.f@gmail.com Dr. Mojgan Sheikhpour, Department of Mycobacteriology and Pulmonary Research, Pasteur Institute of Iran. Tehran, Iran. Microbiology Research Center, Pasteur Institute of Iran, Tehran, Iran. E-mail: mshaikhpoor@gmail. com



This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

that the expression of mir-125b is decreased TB and NSCLC.^[12,13] The results of other studies showed that the function of mir-125b in NSCLC is in the apoptosis pathway,^[14] proliferation, invasion, and migration of NSCLC cells.^[15,16] In addition, TB affects the nature of the immune response.^[13] The research results have shown that mir-125b can be used as a prognostic biomarker in NSCLC and TB.^[17] Since the results of previous studies on the blood and tissue samples in NSCLC patients have shown different results in terms of an alteration of miR125b expression profile in different stages of the disease, in this research study, the expression pattern of miR125b in plasma samples of new-case patients was been examined.

Materials and Methods

This research has been approved by the research committee of Biological Science School, Varamin-Pishva Branch, Islamic Azad University, and confirmed in the Ethics Committee of Pasteur Institute of Iran. Furthermore, a written informed consent form was obtained from participants.

The study population included thirty healthy controls, thirty TB new case patients, and 30 new NSCLC patients. All of the participants were between the ages of 25 and 55 years, before treatment and without any history of surgery. The characteristics of the patients of the present study are summarized in Table 1.

After sampling peripheral blood from the patient's cubital vein, the samples were kept in the tubes containing an anticoagulant (ethylenediaminetetraacetic acid). Then, the samples were centrifuged with a speed of 7000 g for 10 min to separate the plasma. *Total RNA was then isolated* from plasma by the RNA blood kit (Roche, Germany). The quality of RNA was determined using the NanoDrop (Thermo scientific).

The cDNA was synthesized by Transcriptor First Strand cDNA Synthesis Kit (Roche, No. 04 379 012 001,

Germany). To transcription the miRNAs into the first-strand cDNA, the stem-loop primer was used. The stem-loop primer binds to the 3' end of the miRNA and lengths the cDNA,^[18] as shown in Figure 1.

The expression pattern of mir-16 as a reference gene and mir-125b was analyzed in TB patients, NSCLC patients and healthy controls by real-time polymerase chain reaction (PCR) in Rotor-Gene 6000 (Corbett Research, Australia). The LightCycler FastStart DNA Master PLUS Kit SYBR Green I (Roche, Germany) was used for real-time PCR. Primer sequences were designed by the primer express software to exclude amplification of mir-125b and the reference gene mir-16 [Table 2].

Statistical analysis

Data were analyzed by SPSS (version 20) and Graph Pad Prism 6 Software (San Diego, California, USA). Therefore, the fold change and P value were calculated to determine the expression changes of mir-125b in TB, NSCLC Patients

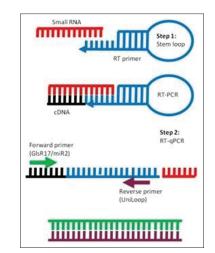


Figure 1: Stem-loop quantitative reverse transcription reverse transcription-quantitative polymerase chain reaction design to identify miRNAs^[19]

control								
Patients	Age	Hepatitis	HIV	Family history	Family history	NonIranian	Smoking	Sex (male/
group	(years)	(%)	(%)	of TB (%)	of NSCLC (%)	(%)	(%)	female) (%)
ТВ	25-55	13	15	30	25	35	35	47/53
NSCLC	25-55	5	5	15	10	10	70	95/5
TD T 1	1		11.1					

TB: Tuberculosis, NSCLC: Nonsmall cell lung cancer

Table 2: The 5' \rightarrow 3 sequences of micro RNAs specific primers							
miRNA name	Sequence $(5' \rightarrow 3')$						
miRNA-16 forward	CCGGAGTAGCAGCACGTAAAT						
miRNA-16 reverse	ATCCAGTGCAGGGTCCGA						
miRNA-16 stem lope	GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACCGCCAA						
miRNA-125b forward	CCTAGATTCCCTGAGACCCT						
miRNA-125b reverse	ATCCAGTGCAGGGTCCGA						
miRNA-125b stem lope	GTCGTATCCAGTGCAGGGTCCGAGGTATTCGCACTGGATACGACTCACAA						

compared to healthy controls. Analysis of variance test was used for comparison of patients groups.

Results

The statistical analysis was performed to calculate the expression pattern of mir-125b in NSCLC (n = 30), TB (n = 30), and normal (n = 30) subjects. According to Figure 2, the expression levels of plasma miR-125b were significantly different in TB and NSCLC patients than in healthy controls.

Statistical analysis using SPSS software (Version 20) (San Diego-California,USA) showed that the expression level of miR-125b in the patients with NSCLC was significantly lower than that in the control subjects (P < 0.0001). Furthermore, miR-125b showed significantly lower expression in patients with TB than in the control individuals (P < 0.0001). The results also showed that the expression changes of mir-125 were significantly similar in TB and NSCLC diseases, which showed that miR-125b had considerable prognostic power to the screening of NSCLC and TB.

Discussion

Cancer is one of the major health problems and the leading cause of death in developed countries and accounts for a high percentage of deaths worldwide.^[20] Exclusively, lung cancer is the most common cause of cancer-related deaths among men and women.^[21] TB is one of the most important infectious diseases of the century that can affect

all body organs, but the lungs are more likely to develop TB.[22] MiRNAs are involved in various physiological and developmental processes such as insulin secretion, apoptosis, hematopoiesis, brain morphogenesis, or tissue differentiation. They are also involved in the immune system and viral diseases.^[23-25] Several types of research have been done to investigate the expression profile pattern of different miRNAs in lung cancer and TB.^[26] In a study conducted in 2016, the diagnostic value of 29 microRNAs for the diagnosis of TB in children was investigated. Fifteen cases of microRNAs were overexpressed, and 14 cases of microRNAs were downregulated. MiR-1, miR-155, miR 31, miR 146a, miR 10a, miR 125b, and miR 150 have decreased expression and miR 29 has increased expression.^[27] The present study results also show that miR-125 has a reduced expression in TB patients, which is in agreement with this study.

By reviewing the KEGG database, we found that the microRNAs have a critical regulatory pathway in lung cancer cancers. Decreasing and increasing the expression of each can have a significant impact on tumorigenesis, angiogenesis, and metastasis. That way, as shown in Figure 3, the downregulation of miR-125b leads to metastasis.

On the other hand, other studies on lung cancer and TB have also been investigated in other miRNAs and reported as diagnostic biomarkers. For example, in 2016, a study by Zhu *et al.* On altered miRNA expression of miR-182, miR-183, and miR-210, miR-126, in smokers, pneumonia

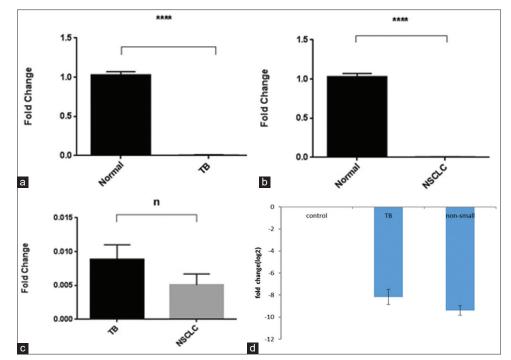


Figure 2: MiRNA-125 expression patterns in both tuberculosis and nonsmall-cell lung cancer patients are significantly lower than normal individual (a) tuberculosis and normal, (b) nonsmall-cell lung cancer and normal (c) tuberculosis and nonsmall-cell lung cancer, and (d) tuberculosis and nonsmall-cell lung cancer in comparison with normal individuals

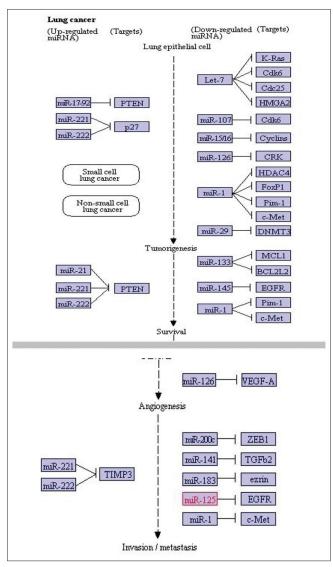


Figure 3: The data obtained from Kyoto Encyclopedia of Genes and Genomes from the roles of Micro RNAs in lung cancer

patients, people with NSCLC, people with gastric cancer, and normal individuals were studied, and the results of their research showed that these four miRNAs could be used as a diagnostic biomarker for lung cancer.^[28] However, some researchers reported that the mir 125 b has a high expression level in NSCLC tissues. The results of a study by Wang *et al.* in 2017 on the expression of MiR-125b in NSCLC showed that miR-125b expression in NSCLC tissue was higher than that in paracarcinoma tissue.^[29]

In comparison, our study was done on plasma sample sources of TB and NSCLC patients. Besides, Yuxia *et al.* analyzed serum levels of miR-125b in 193 patients with different stages of NSCLC in 2012. They found that serum miR-125b was consistently expressed in the nontumor group and was significantly associated with the NSCLC stage, and also patients with high miR-125b expression displayed a substantially poorer prognosis compared with patients with low expression (P < 0.0001).^[17]

Studies have shown that it has been found that markers for lung cancer and TB have been found so far. In the present study, miR-125b is described as a prognostic even may be diagnostic biomarker in lung cancer and TB, which must be checked in a large number of patients in larger study groups. Furthermore, the expression profile if mir 125 b should be evaluated at different stages of these diseases in both the blood and tissue samples. Be more accurately assessed to have a diagnostic value. On the other hand, several studies have shown that TB and lung cancer overlap in many cases and TB is a precursor to lung cancer. Hence, mir 125 b could be used for screening of TB and NSCLC patients. It has been often said that TB can be a risk factor for lung cancer. People with TB usually have drug resistance and in many cases, develop lung cancer, so this biomarker can help follow-up on the TB patient to prevent getting lung cancer or lung cancer patient from the risk of metastasis.

Acknowledgment

We thank all of the patients and healthy individuals who contributed and helped us in this research by donating their blood samples. Furthermore, we thank our colleagues from the Department of Mycobacteriology and Pulmonary Research, Microbiology Research Center, Pasteur Institute of Iran. The authors acknowledge the Oncology Department of Masih Daneshvari Hospital, Dr. Adnan Khosravi, to arrange the sampling process.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

- 1. Zappa C, Mousa SA. Non-small cell lung cancer: Current treatment and future advances. Transl Lung Cancer Res 2016;5:288-300.
- Ishida M, Iwai M, Kagotani A, Iwamoto N, Okabe H. Cutaneous metastasis from pulmonary large cell neuroendocrine carcinoma in the scalp. Int J Clin Exp Pathol 2014;7:2701-6.
- Farhat MR, Freschi L, Calderon R, Ioerger T, Snyder M, Meehan CJ, et al. GWAS for quantitative resistance phenotypes in *Mycobacterium tuberculosis* reveals resistance genes and regulatory regions. Nat Commun 2019;10:2128.
- Baumjohann D, Ansel KM. MicroRNA-mediated regulation of T helper cell differentiation and plasticity. Nat Rev Immunol 2013;13:666.
- Rusek AM, Abba M, Eljaszewicz A, Moniuszko M, Niklinski J, Allgayer H. MicroRNA modulators of epigenetic regulation, the tumor microenvironment and the immune system in lung cancer. Mol Cancer 2015;14:34.
- Meola N, Gennarino VA, Banfi S. MicroRNAs and genetic diseases. Pathogenetics 2009;2:7.
- Tsai LM, Yu D. MicroRNAs in common diseases and potential therapeutic applications. Clin Exp Pharmacol Physiol 2010;37:102-7.

- Cho WC. MicroRNAs in cancer From research to therapy. Biochim Biophys Acta 2010;1805:209-17.
- 9. Ruan K, Fang X, Ouyang G. MicroRNAs: Novel regulators in the hallmarks of human cancer. Cancer Lett 2009;285:116-26.
- O'Connell RM, Rao DS, Chaudhuri AA, Baltimore D. Physiological and pathological roles for microRNAs in the immune system. Nat Rev Immunol 2010;10:111-22.
- 11. Tsitsiou E, Lindsay MA. MicroRNAs and the immune response. Curr Opin Pharmacol 2009;9:514-20.
- Yu X, Wei F, Yu J, Zhao H, Jia L, Ye Y, *et al.* Matrix metalloproteinase 13: A potential intermediate between low expression of microRNA-125b and increasing metastatic potential of non-small cell lung cancer. Cancer Genet 2015;208:76-84.
- Rajaram MV, Ni B, Morris JD, Brooks MN, Carlson TK, Bakthavachalu B, *et al. Mycobacterium tuberculosis* lipomannan blocks TNF biosynthesis by regulating macrophage MAPK-activated protein kinase 2 (MK2) and microRNA miR-125b. Proc Natl Acad Sci U S A 2011;108:17408-13.
- Biamonte F, Battaglia AM, Zolea F, Oliveira DM, Aversa I, Santamaria G, *et al.* Ferritin heavy subunit enhances apoptosis of non-small cell lung cancer cells through modulation of miR-125b/p53 axis. Cell Death Dis 2018;9:1174.
- Li Y, Chao Y, Fang Y, Wang J, Wang M, Zhang H, *et al.* MTA1 promotes the invasion and migration of non-small cell lung cancer cells by downregulating miR-125b. J Exp Clin Cancer Res 2013;32:33.
- Li X, Zhang Z, Jiang H, Li Q, Wang R, Pan H, *et al.* Circular RNA circPVT1 promotes proliferation and invasion through sponging miR-125b and activating E2F2 signaling in non-small cell lung cancer. Cell Physiol Biochem 2018;51:2324-40.
- Yuxia M, Zhennan T, Wei Z. Circulating miR-125b is a novel biomarker for screening non-small-cell lung cancer and predicts poor prognosis. J Cancer Res Clin Oncol 2012;138:2045-50.
- Kramer MF. Stem-loop RT-qPCR for miRNAs. Curr Protoc Mol Biol 2011;95:1-15.
- 19. Marcial-Quino J, Gómez-Manzo S, Fierro F, Vanoye-Carlo A,

Rufino-González Y, Sierra-Palacios E, *et al.* Stem-Loop RT-qPCR as an Efficient Tool for the Detection and Quantification of Small RNAs in Giardia lamblia. Genes (Basel) 2016;7:131.

- Rafiquzzaman R. Cytotoxic effects of quercus infectoria extracts towards cervical (Hela) and ovarian (Caov-3) cancer cell lines. Health Environ J 2010;1.
- DeSantis C, Naishadham D, Jemal A. Cancer statistics for African Americans, 2013. CA Cancer J Clin 2013;63:151-66.
- 22. Mustafa AD, Kalyanasundram J, Sabidi S, Song AA, Abdullah M, Abdul Rahim R, *et al.* Recovery of recombinant *Mycobacterium tuberculosis* antigens fused with cell wall-anchoring motif (LysM) from inclusion bodies using non-denaturing reagent (N-laurylsarcosine). BMC Biotechnol 2019;19:27.
- Büssing I, Slack FJ, Grosshans H. Let-7 microRNAs in development, stem cells and cancer. Trends Mol Med 2008;14:400-9.
- 24. Cho WC. MicroRNAs in cancer-From research to therapy. Biochim Biophys Acta 2010;1805:209-17.
- 25. Giovannetti E, Erozenci A, Smit J, Danesi R, Peters GJ. Molecular mechanisms underlying the role of microRNAs (miRNAs) in anticancer drug resistance and implications for clinical practice. Crit Rev Oncol Hematol 2012;81:103-22.
- 26. Abolfathi H, Sheikhpour M, Mohammad Soltani B, Fahimi H. The Comparison and Evaluation of the miR-16, miR-155 and miR-146a Expression Pattern in the Blood of TB and NSCLC Patients: A Research Paper. Gene Reports. 22. 2021. p. 100967.
- Zhou M, Yu G, Yang X, Zhu C, Zhang Z, Zhan X. Circulating microRNAs as biomarkers for the early diagnosis of childhood tuberculosis infection. Mol Med Rep 2016;13:4620-6.
- Zhu W, Zhou K, Zha Y, Chen D, He J, Ma H, *et al.* Diagnostic value of serum miR-182, miR-183, miR-210, and miR-126 levels in patients with early-stage non-small cell lung cancer. PLoS One 2016;11:e0153046.
- Wang Y, Zhao M, Liu J, Sun Z, Ni J, Liu H. miRNA-125b regulates apoptosis of human non-small cell lung cancer via the PI3K/Akt/GSK3β signaling pathway. Oncol Rep 2017;38:1715-23.